

PHYSIOLOGICAL EFFECTS OF DIFFERENT TYPES OF β -GLUCAN

Vaclav Vetvicka*, Jana Vetvickova

University of Louisville, Department of Pathology, Louisville, KY, USA
e-mail: Vaclav.vetvicka@louisville.edu

Received: August 22, 2007; Accepted: October 5, 2007

Key words: Glucan/Phagocytosis/Cholesterol/Blood sugar

Background: Numerous types of glucans have been isolated from almost every species of yeast, grain, and fungi. These products have been extensively studied for their immunological and pharmacological effects.

Aim: In this paper we evaluated the possibility whether individual glucans will be similarly active against each of the tested biological properties or if each glucan will affect different reactions.

Methods: Immunological effects of glucans were measured by evaluation of phagocytosis of HEMA particles by peripheral blood leukocytes and production of IL-2 by mouse splenocytes. Next we measured the effects of long-term treatment with glucan on levels of blood glucose and blood cholesterol. Four different glucans differing in origin (yeast, grain and mushroom) were used.

Results: Our results showed that the same glucan, yeast-derived insoluble #300 glucan, stimulated phagocytosis of peripheral blood leukocytes, production of IL-2 by mouse splenocytes, lowered the cholesterol levels in mice with experimentally-induced cholesterolemia and lowered the level of blood sugar after induced hyperglycaemia. The remainder of tested glucans were only marginally active.

Conclusion: Taken together, our study showed that with respect to natural glucans, there is a yes-or-no effect suggesting that highly purified and highly active glucans will have pleiotropic impact, whereas poorly isolated and/or less active glucans will have only mediocre biological properties.

INTRODUCTION

Natural products, useful in treating or preventing various diseases, have been sought throughout the history of mankind. Most of these natural products are plagued with a common problem, i.e., the fact that they often represent a complex mixture of individual ingredients each of which can contribute to their biological activity. Natural (1,3)-beta-D-glucans from yeast, grain and mushrooms are well-established biological response modifiers^{1,2}, representing highly conserved structural components of cell walls in yeast, fungi, seaweed or grain seeds. 1,3-Glucan's role as a biologically active immunomodulator has been well documented for over 40 years. Interest in the immunomodulatory properties of polysaccharides was initially raised after experiments showing that a crude yeast cell preparation stimulated macrophages via activation of the complement system³.

Numerous types of glucans have been isolated from almost every species of yeast, grain and fungi. (1,3)-beta-D-glucans have been extensively studied for their immunological and pharmacological effects. More than 2,000 papers describing the mostly immunological activities of glucans exist in the literature (for review see⁴).

However, multiple reports established additional effects of glucans. Nearly 50 years ago the possible effects of dietary fiber were first suggested by Keys⁵ and these effects were later found to be associated with glucans⁶. The cholesterol-lowering effects of fibers are routinely associated

with beta-glucans. Due to the high consumption of oats or oat bran, most of the attention has been focused on the relationship of oat-derived glucan to cholesterol levels. These glucans were shown to reduce serum cholesterol in both hypercholesterolemic animals⁷⁻⁹ and humans¹⁰, however there are some studies that found no such effects¹¹.

In addition, fiber-containing glucans, particularly the ones in human food, gained attention for their role in the metabolic control of diabetes¹²⁻¹⁵. However, most of these studies were accomplished using only partly purified glucans, making evaluation of the glucan role rather difficult.

Despite extensive investigations, no consensus on the source, size and other biochemical or physicochemical properties of beta-1,3-glucan has been achieved. In addition, numerous concentrations and routes of administration have been tested - including oral, intraperitoneal, subcutaneous and intravenous applications. Besides, there are probably more than a hundred of different samples on the U.S. market alone, which leads to a confusion about the quality, biological effects and overall efficiency of glucan. Therefore, we decided to compare the basic biological and immunological activities of a group of glucans.

The collection of techniques including phagocytosis and stimulation of IL-2 synthesis effects on cholesterol and blood sugar levels represents a general approach to the biological characteristics of tested glucans thus offering insight as to their biological activities.

Table 1.

	Control	#300	Krestin	Immunofibre	WGP	Now Yeast
Total weight	23.33 ± 1.06	24.34 ± 1.27	24.11 ± 1.48	24.99 ± 4.11	25.87 ± 3.38	26.99 ± 2.77
Liver	1.77 ± 0.23	1.49 ± 0.31	1.55 ± 0.19	1.54 ± 0.38	1.66 ± 0.35	1.47 ± 0.35
Spleen	0.16 ± 0.09	0.17 ± 0.06	0.14 ± 0.05	0.17 ± 0.07	0.15 ± 0.04	0.16 ± 0.03
Thymus	0.05 ± 0.02	0.06 ± 0.01	0.06 ± 0.01	0.05 ± 0.01	0.05 ± 0.05	0.08 ± 0.02
Heart	0.16 ± 0.06	0.17 ± 0.10	0.18 ± 0.08	0.16 ± 0.03	0.16 ± 0.05	0.17 ± 0.03
Kidneys	0.41 ± 0.10	0.38 ± 0.07	0.40 ± 0.12	0.40 ± 0.15	0.36 ± 0.08	0.42 ± 0.05
Lung	0.16 ± 0.02	0.17 ± 0.02	0.15 ± 0.02	0.18 ± 0.03	0.16 ± 0.04	0.18 ± 0.05

MATERIAL AND METHODS

Animals. Female, 8 weeks old BALB/c mice were purchased from the Jackson Laboratory (Bar Harbor, ME). All animal work was done according to the University of Louisville IACUC protocol. Animals were sacrificed by CO₂ asphyxiation.

Materials. RPMI 1640 medium, sodium citrate, Wright stain, Limulus lysate test E-TOXATE, Concanavalin A, cholesterol, streptozocin and were obtained from Sigma Chemical Co. (St. Louis, MO). Fetal calf serum (FCS) was from Hyclone Laboratories (Logan, UT).

Diet. All diets (Laboratory Rodent Diet 5001 enhanced with glucan and/or cholesterol) were formulated and prepared by Purina (Richmond, IN). Diet ingredients for all groups were identical except for the proportion of glucan and/or cholesterol. Glucan-enhanced diet contained various doses of individual glucan, cholesterol-enhanced diet contained cholesterol. The beta-glucans were quantified by using a Megazyme kit.

Beta-1,3 glucans. Yeast-derived insoluble glucan #300 were purchased from Transfer Point (Columbia, SC), mushroom-derived Krestin from (Kureha Chemical Industries, Tokyo, Japan), yeast-derived Now BETA glucan from Now Foods (Bloomington, IL), and grain-derived ImmunoFiber was purchased from (Whole Control, Arvada, CO).

Phagocytosis. The technique employing phagocytosis of synthetic polymeric microspheres was described earlier^{15,16}. Briefly: peripheral blood cells were incubated with 0.05 ml of 2-hydroxyethyl methacrylate particles (HEMA; 5 × 10⁸/ml). The test tubes were incubated at 37 °C for 60 min., with intermittent shaking. Smears were stained with Wright stain. The cells with three or more HEMA particles were considered positive. The same smears were also used for evaluation of cell types.

Evaluation of IL-2 production. Purified spleen cells (2 × 10⁶/ml in RPMI 1640 medium with 5% FCS) were added into wells of a 24-well tissue culture plate. After addition of 1 µg of Concanavalin A into positive-control wells, cells were incubated for 72 hrs in a humidified incubator (37 °C, 5% CO₂). At the endpoint of incubation, supernatants were collected, filtered through 0.45 µm filters and tested for the presence of IL-2. Levels of the IL-2 were measured using a Quantikine mouse IL-2 kit (R&D Systems, Minneapolis, MN).

Glucose evaluation. The mice were given drinking water freely and were not fed 24 hr prior to measurement of blood glucose level. In some experiments, blood glucose was determined in hyperglycaemic mice which were pre-treated with streptozotocin (250 mg/kg ip.) 12 days before the start of feeding with glucan¹⁷.

Biochemical Analysis. Mice were deprived of food for 24 hr and sacrificed. Serum was collected via the retro-orbital sinus and stored at - 80 °C for less than a week. Biochemical analyses were performed by Antech Diagnostics (Indianapolis, IN).

Statistics. Student's t-test was used to statistically analyze the data.

RESULTS

Phagocytosis is one of the biological activities traditionally connected with effects of natural immunomodulators, including glucans. Therefore, we initiated our study by comparison of the effects of intraperitoneally applied glucans used as a single dose (Fig. 1). From day 1, glucan #300 showed much stronger activation of blood neutrophils than other glucans. Two other glucans, Krestin and NOW glucan, also showed significant effects lasting up to 48 hrs. When we repeated the glucan administration for three consecutive days, we found not only higher phagocytic activity but also that it lasted significantly longer - in the case of #300 and Krestin up to 7 days (data not shown).

Production of IL-2 belongs to the valuable indicators of the immune activities. Therefore, we compared the effects of tested glucans on the secretion of IL-2 by spleen cells isolated from glucan-treated mice. The IL-2 production was measured after a 72 hr *in vitro* incubation of cells. The results summarized in Fig. 2 showed that even when all tested glucan stimulated IL-2 production, there were huge differences between individual glucans (i.e., #300 stimulated IL-2 secretion 3.5 times more than ImmunoFiber). The activity of all tested glucans slowly decreased with time but was still measurable 14 days after injection.

For cholesterol and blood sugar experiments we used the Laboratory Rodent Diet 5001 consisting of 23.9 % protein, 4.6 % fat, 5.5 % fiber, with 75.5 % total digestible nutrients. This diet was supplemented with either glucan or cholesterol corresponding to the final daily doses of

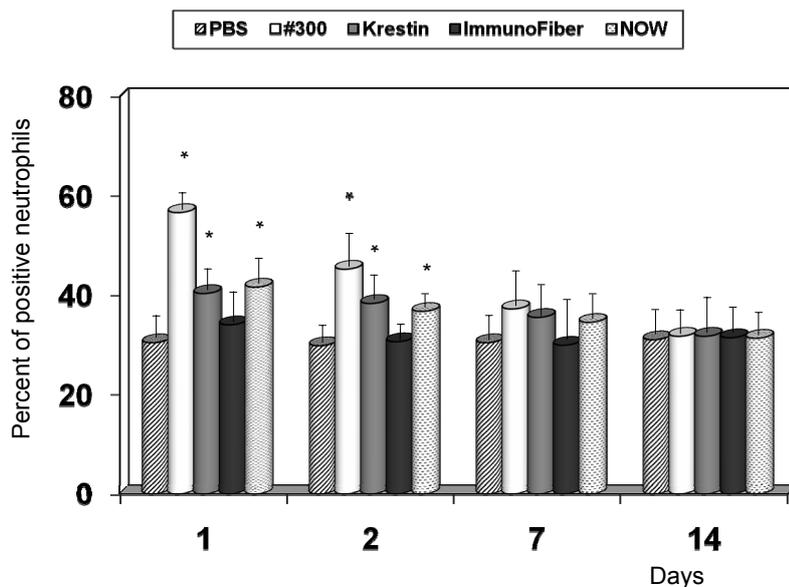


Fig. 1. Effect of long-term feeding with individual glucans (see Materials and Methods) on phagocytosis of HEMA particles by peripheral blood neutrophils. Each value represents the mean of three independent experiments \pm SD.

*Represents significant differences between control (PBS) and glucan samples at $P < 0.05$ level.

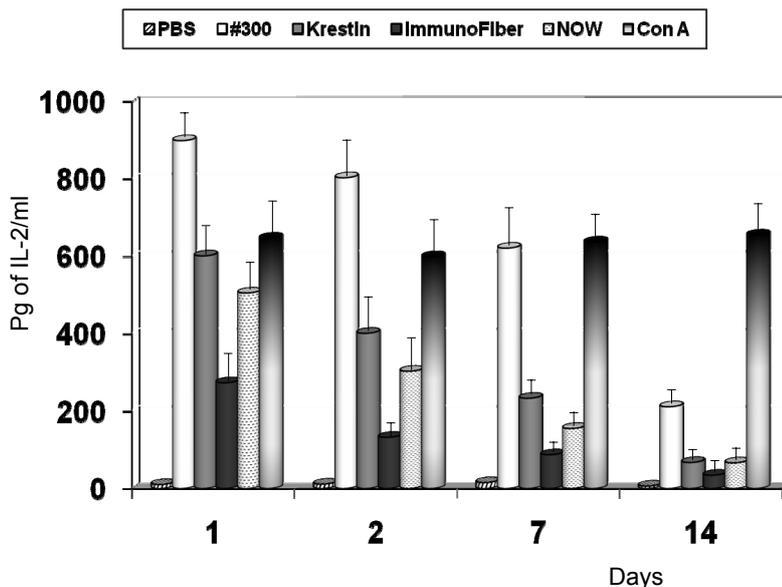


Fig. 2. Effects of glucan feeding on Con A-stimulated secretion of IL-2 by spleen cells.

*Represents significant differences between control (PBS) and samples at $P < 0.05$ level.

100 μ g of glucan or 16 μ g of cholesterol, respectively. The mice adapted well to all diets and all survived the dietary treatment. Body weights and individual organ weights did not differ among the groups at each time point (Table 1).

First, we studied the effect of long-term feeding with glucan-added diet. Our data showed strong time-dependent effects of #300 and Krestin glucans on lowering cholesterol. Effects of other glucans were less pronounced and in the case of ImmunoFiber we found almost no effects (except after 50 days of feeding) - (Fig. 3)

The mice were then given a diet with added cholesterol. The blood cholesterol levels obtained after two weeks of cholesterol feeding were used as positive control. The cholesterol-rich diet was followed by 40 days of feeding with glucan-rich diet. Individual groups of mice were sacrificed in 10-day-intervals and cholesterol levels were evaluated. Results summarized in Fig. 4 showed that during short time intervals all glucans lowered the cholesterol levels in hypercholesterolemic animals, but in the long term only #300 glucan retained this activity.

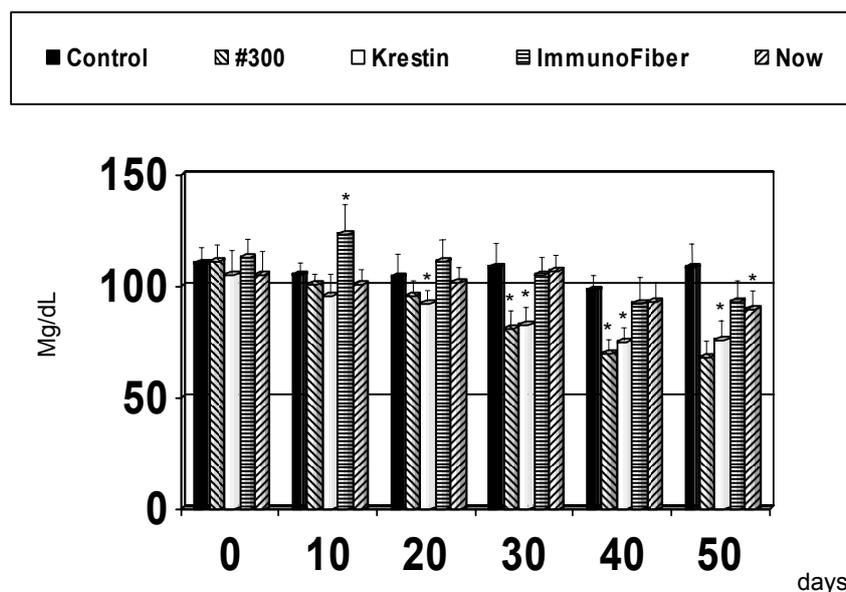


Fig. 3. Effect of long-term feeding with individual glucans (see MATERIALS AND METHODS) on blood cholesterol levels. Each value represents the mean of three independent experiments \pm SD.

*Represents significant differences between control (PBS) and glucan samples at $P \leq 0.05$ level.

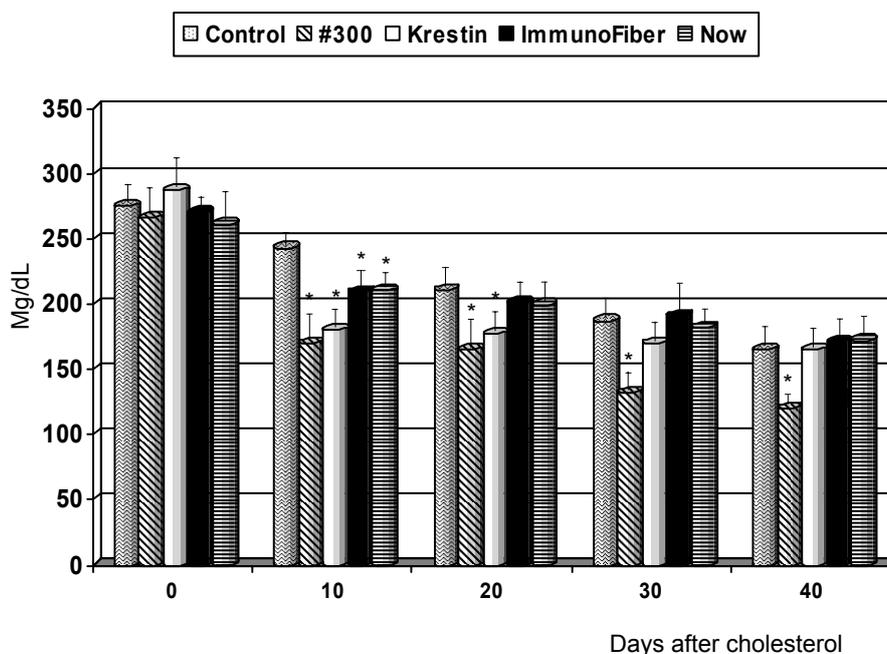


Fig. 4. Effect of long-term feeding with individual glucans on blood cholesterol levels in experimentally-induced hypercholesterolemia. The feeding with glucan started after two weeks of cholesterol-high diet (see Materials and Methods). Control group (Control) was fed with standard diet. Each value represents the mean of three independent experiments \pm SD.

*Represents significant differences between control (PBS) and glucan samples at $P \leq 0.05$ level.

The final part of our project was focused on effects of glucan administration on levels of blood sugar. Feeding with glucan did not significantly affect the sugar levels (Fig. 5). However, a different situation was found when we used mice with experimentally induced hyperglycaemia. After two weeks of feeding, #300 glucan significantly lowered the sugar levels to almost a normal (compare Figures 5 and 6). A longer application of glucan resulted in additional significant activity of ImmunoFibre glucan.

DISCUSSION

β -Glucans show notable physiological effects. This is the main reason they are of such great interest. They belong to a group of physiologically active compounds, collectively termed biological response modifiers. Thus far, among many known and tested immunomodulators of the first order, polysaccharides isolated from different microorganisms and plants hold a formidable place.

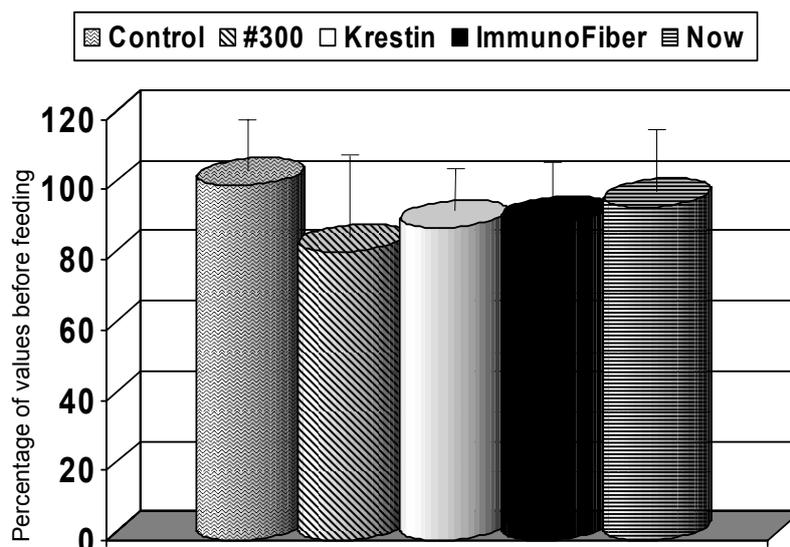


Fig. 5. Effect of long-term feeding with individual glucans (see MATERIALS AND METHODS) on blood glucose levels. The results are given as percentage of control values. Each value represents the mean of three independent experiments \pm SD.

*Represents significant differences between control (PBS) and glucan samples at $P \leq 0.05$ level.

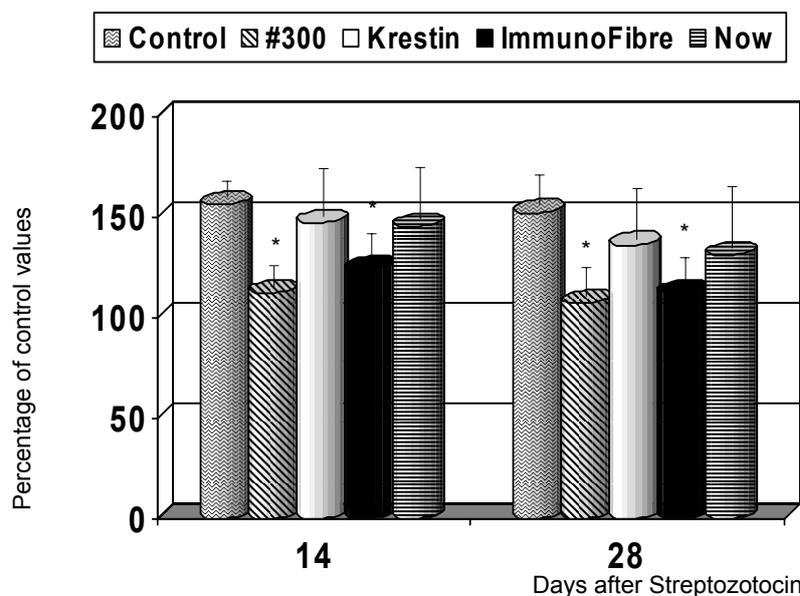


Fig. 6. Effect of long-term feeding with individual glucans (see MATERIALS AND METHODS) on blood glucose levels in mice with experimentally-induced hyperglycaemia. The results are given as percentage of control values. Each value represents the mean of three independent experiments \pm SD.

*Represents significant differences between control (PBS) and glucan samples at $P \leq 0.05$ level.

A large number of such polysaccharides, that act only as immunopotentiators, are well known¹⁸.

Binding of β -glucan to specific receptors (either CR3 or Dectin-1) activates macrophages. The activation consists of several interconnected processes including increased chemokinesis, chemotaxis, migration of macrophages, degranulation leading to increased expression of adhesive molecules, and adhesion to the endothelium. In addition, β -glucan binding triggers intracellular processes, characterized by the respiratory burst after phagocytosis of invading cells (formation of reactive oxygen species

and free radicals), the increase of content and activity of hydrolytic enzymes, and signaling processes leading to activation of other cells and secretion of cytokines. For an excellent review regarding interaction of glucans with macrophages see¹⁹.

The rationale for the choice of glucans parallels what was explained in our previous paper²⁰. We chose four glucans widely sold and available in the US, Europe and the Far East, representing grain-, mushroom- and yeast-derived glucans in soluble and insoluble form. Briefly, #300 is insoluble yeast-derived glucan; Krestin is soluble

mushroom-derived glucan; ImmunoFiber represents soluble grain-derived glucan; and NOW is a mixture of both insoluble glucans from yeast and soluble glucans from mushrooms.

There are very few comprehensive reviews focused on biological properties of glucans from various sources that exist. The comparative reviews focus mainly toward the reflection of chemical characteristics of glucans on their biological and immunological properties^{21, 22}.

In this paper, we continued the comparison of several commercially important glucans. Glucans are well known for their ability to stimulate the innate immunity and the cellular branch of immune reaction¹⁸. Therefore, our initial focus was phagocytic activity with the use of peripheral blood neutrophils and synthetic microspheres as a model. Our results confirmed our previous studies showing that glucan #300 was one of the most active glucans available.

It is assumed that glucan application results in signaling processes leading to activation of macrophages and other cells and subsequent secretion of cytokines and other substances initiating inflammation reactions (e.g., interleukins IL-1, IL-2, IL-6, TNF- α)²³⁻²⁵. We found that all tested glucans stimulated splenocytes to produce IL-2, with #300 and Krestin showing the strongest and longest effects.

Most of experimental studies dealing with cholesterol-lowering effects of glucans used oats- or barley-derived glucans^{8, 9}, without finding significant differences between these glucans⁷. The cholesterol-lowering effects of glucans are well-established on numerous models, including humans^{8, 26}. Without any direct proof, these effects are usually described as the result of fiber intake and subsequent decreased absorption of bile acids²⁷. However, most of these studies suffer from the fact that they did not evaluate the effects of isolated glucans. They only used crude extracts²⁸ without any knowledge if these glucans are even digested.

The current study is not only the first to directly compare the cholesterol-lowering activity of two different yeast-derived β -glucans, but also the first to compare normal animals and mice with experimentally induced cholesterolemia.

The effects of glucans on blood sugar levels are less known. Some studies showed hypoglycemic activity of natural glucans^{13, 29}, additional studies demonstrated strong hypoglycemic activity of synthetic polysaccharides^{17, 30}. The mechanisms remain, however, unknown. Some groups suggested similar mechanisms as in lowering cholesterol, i.e., changes in the increase of viscosity of the alimentary bolus and changes in the gastric emptying³¹. Our data found no correlation between these effects and source of the glucan or its solubility.

The present paper represents yet additional proof of the vast differences among commercially available glucans. Based on the presented data, it is clear that the individual glucans are either active in all tested areas or their activity is only mediocre. It is most likely that this yes-or-no effect suggests there are no glucans that can be tailor-made for stimulation of one particular reaction.

ACKNOWLEDGEMENT

Authors of this study have no financial interest in any of the products or manufacturers mentioned in this article. No external funding was provided for this study. The authors would like to thank Ms. Rosemary Williams for excellent editorial assistance.

REFERENCES

1. Borchers AT, Stern JS, Hackman RM, Keen CL, Gershwin ME. Mushrooms, tumors, and immunity. *Proc Soc Exp Biol Med* 1999; 221:281-293.
2. Brown GD, Gordon S. Fungal β -glucans and mammalian immunity. *Immunity* 2003; 19:311-315.
3. Benacerraf B, Sebastyen MM. Effect of bacterial endotoxins on the reticuloendothelial system. *Fed Proc* 1957; 16: 860-867.
4. Novak M, Větvická V. Beta-glucans, history and present. *Alternative Medicine Review* 2007, in press.
5. Keys A, Anderson JT, Grande F: Diet-type (fast constant) and blood lipid in man. *J Nutr* 1960; 70:257-266.
6. Tietzen JL, Nevins DJ, Schneeman BO: Characterization of the hypocholesterolemic potential of oat bran. *FASEB J* 1990; 4: A527.
7. Delaney B, Nicolosi RJ, Wilson TA, Carlson T, Frazer S, Zheng GH, et al. Beta-glucan fractions from barley and oats are similarly antiatherogenic in hypercholesterolemic Syrian golden hamsters. *J Nutr* 2003; 133:468-475.
8. Fadel JG, Newman RK, Newman CW, Barnes AE: Hypocholesterolemic effects of beta-glucans in different barley diets fed to broiler chicks. *Nutr Rep Int* 1987; 35:1049-1058.
9. Kahlon TS, Chow FI, Knuckles BE, Chiu MM. Cholesterol-lowering effects in hamsters of beta-glucan-enriched barley fraction, dehulled whole barley, rice bran, and oat bran and their combinations. *Cereal Chem* 1993; 70: 435-440.
10. Queenan KM, Stewart ML, Smith KN, Thomas W, Fulcher RG, Slavin JL. Concentrated oat beta-glucan, a fermentable fiber, lowers serum cholesterol in hypercholesterolemic adults in a randomized controlled trial. *Nutrition J* 2007; 6:1-8.
11. Keogh GF, Cooper GJ, Mulvey TB, McArdle BH, Coles GD, Monro JA, et al. Randomized controlled crossover study of the effect of a highly beta-glucan-enriched barley on cardiovascular disease risk factors in mildly hypercholesterolemic men. *Am J Clin Nutr* 2003; 78:711-718.
12. Wursch P, Pi-Sunyer FX. The role of viscous soluble fiber in the metabolic control of diabetes: a review with special emphasis on cereals rich in beta-glucan. *Diabetes Care* 1997; 20:1774-1780.
13. De Paula ACCFF, Sousa RV, Figueiredo-Ribeiro, RCL, Buckeridge MS. Hypoglycemic activity of polysaccharide fractions containing beta-glucans from extracts of *Rhynchomytrum repens* (Willd.) C.E. Hubb., *Poaceae*. *Braz J Med Biol Res* 2005; 38:885-893
14. Hong L, Xun M, Wutong W. Anti-diabetic effects of beta-glucan from fruit body of maitake (*Grifola frondosa*) on KK-Ay mice. *J Pharmacy Pharmacol* 2007; 59:575-582
15. Větvická V, Fornusek L, Kopeček J, Kaminková J, Kaspárek L, Vranová M. Phagocytosis of human blood leukocytes: A simple micromethod. *Immunol Lett* 1982; 5:97-100.
16. Větvická V, Holub M, Kováru H, Šiman P, Kováru F. Alpha-fetoprotein and phagocytosis in athymic nude mice. *Immunol Lett* 1988; 19:95-98.
17. Hatanaka K, Song SC, Maruyama A, Kobayashi A, Kuzuhara H, Akaike T. A new synthetic hypoglycaemic polysaccharide. *Biochem Biophys Res Comm* 1992; 188:16-19.
18. Whistler RL, Bushway AA, Singh PP, Nakahara W, Tokuzen R. Noncytotoxic, antitumor polysaccharides. *Adv Carbohydr Chem Biochem* 1976; 32:235-275.
19. Schepetkin IA, Quinn MT. Botanical polysaccharides: macrophage immunomodulation and therapeutic potential. *Int Immunopharmacol* 2006; 6:317-333.

20. Vetvicka V, Vetvickova J. An evaluation of the immunological activities of commercially available beta-1,3-glucans. *JANA* 2007; 10:25-31.
21. Yadomae T. Structure and biological activities of fungal beta-1,3-glucans. *Yakugaku Zasshi* 2000; 120:413-431.
22. Kogan G. (1-3,1-6)-beta-D-glucans of yeast and fungi and their biological activity; in: Atta-ur-Rahman, (ed): *Studies in Natural Products Chemistry*. Amsterdam, Elsevier, 2000, pp 107-152.
23. Abel G, Czop JK. Stimulation of human monocyte beta-glucan receptors by glucan particles induces production of TNF- α and IL-1 β . *Int J Immunopharmacol* 1992; 14:1363-1373.
24. Adachi Y, Okazaki M, Ohno N, Yadomae T. Enhancement of cytokine production by macrophages stimulated with (1->3)-beta-D-glucan, Grifolan (GRN), isolated from *Grifola frondosa*. *Biol Pharm Bull* 1994; 17:1554-1560.
25. Vetvicka V, Terayama K, Mandeville R, Brousseau P, Kournikakis B, Ostroff G. Pilot study: orally-administered yeast beta-1,3-glucan prophylactically protects against anthrax infection and cancer in mice. *JANA* 2002; 5:1-6.
26. Reyna-Villasmil N, Bermudez-Pirela V, Mengual-Moreno E, Arias N, Cano-Ponce C, Leal-Gonzales E, et al. Oat-derived beta-glucan significantly improves HDLC and diminishes LDLC and non-HDL cholesterol in overweight individuals with mild hypercholesterolemia. *Am J Therapeut* 2007; 14:203-212.
27. Malkki Y: Oat fiber; in: Cho SS, Dreher ML, (eds): *Handbook of Dietary Fiber*. New York, Marcel Dekker, 2001, pp 497-512.
28. Wang L, Behr SR, Newman RK, Newman CW: Comparative cholesterol-lowering effects of barley beta-glucan and barley oil in golden syrian hamsters. *Nutr Res* 1997; 17:77-88.
29. Battilana P, Ornstein K, Minehire J, Schwarz JM, Acheson K, Scheniter P, et al., Mechanisms of action of beta-glucan in post-prandial glucose metabolism in healthy men. *Eur J Clin Nutr* 2001; 55:327-333.
30. Sone, Y., Makino, C., Misaki, A. Inhibitory effects of oligosaccharides derived from plant xyloglucan on interstitial glucose absorption in rat. *J Nutr Sci Vitaminol* 1992; 38: 391-395.
31. Wood PJ. Physicochemical properties and physiological effects of the (1-4) (1-4)-beta-D-glucan from oats. *Adv Exp Med Bio.* 1990; 270: 119-127.

